



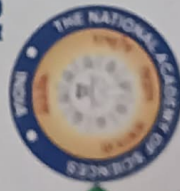
Manoharhal Shikshan Prasarak Mandal, Armori

# RASHTRAPITA MAHATMA GANDHI

ARTS AND SCIENCE COLLEGE NAGBHID DIST - CHANDRAPUR

NAAC Reaccredited 'B+' Grade

In Collaboration With



The National Academy of Science [NASI], India, Nagpur Chapter

## Certificate

This is to certify that, Mr./Mrs. Priyanshu A. Yermadwar  
of Government Science College, Gadchiroli has participated in

### STATE LEVEL STUDENT SEMINAR COMPETITION

On **26<sup>th</sup> February 2024**, in Subject: **Zoology/ Botany/ Chemistry/ Physics**  
And Secured **First/Second/Third/ Consolation Prize/Participated** in event.  
His/Her Valuable Participation in the Competition hereby Acknowledged.

### CONGRATULATIONS

Prof. N.S. Gajbhiye

Chairman

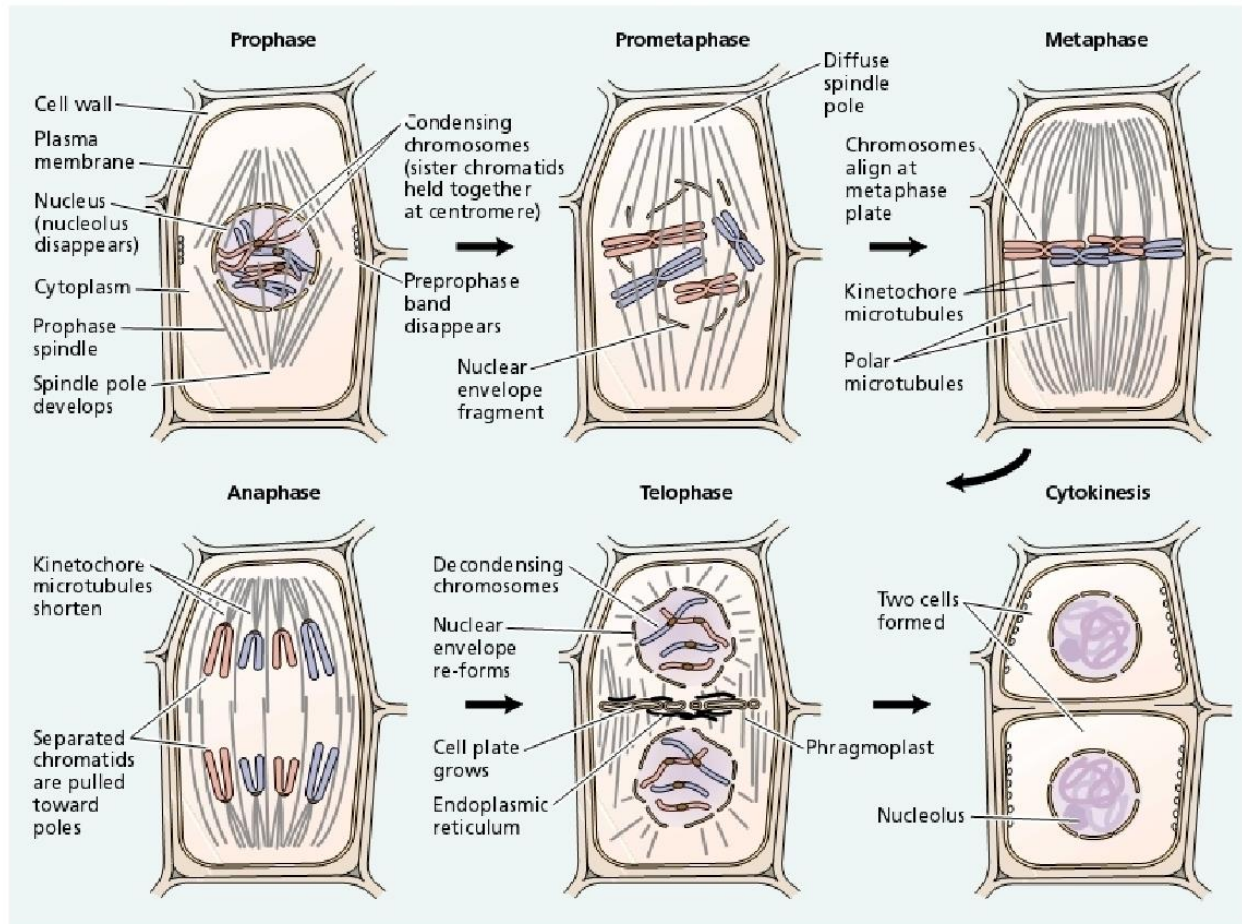
NASI, Nagpur Chapter



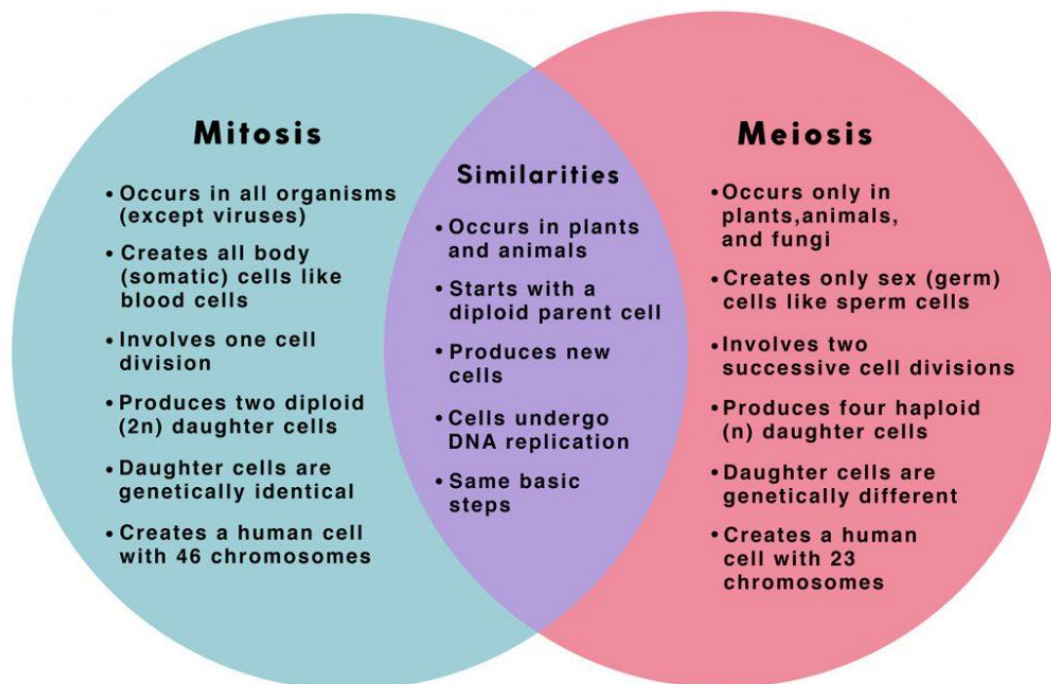
Dr. G.D. Deshmukh

Principal

R.M.G. College, Nagbhid



## Mitosis and Meiosis Venn Diagram



**GOVT. SCIENCE COLLEGE GADCHIROLI**  
**PG DEPARTMENT OF BOTANY**  
**LIST OF PROJECTS CARRIED OUT BY STUDENTS**  
**M.Sc II (Botany) (CBCS), Sem IV**  
**Session : 2023-24**

Sr. No	Name of Student	Title of Project	Type Lab Work /Field Work, Survey/ Other	Name of Supervisor
1.	Miss. Rabina Samar Mandal	“Wild edible vegetables in chamorshi”	Field Work, Survey	Dr. P. S. Jakhi
2.	Miss. Anjali Nandkishor Sonkusare	“Effects Of Organic Manures On Seed Germination in Some Vegetable Crops”	Lab Work	Dr. Shagufta. A. Sheikh
3.	Miss. Punam Anandrao Nagpurkar	“Influence of organic manure on seed germination in some vegetable crops”	Lab Work	Dr. Shagufta A. Sheikh
4.	Miss. Maheshwari Vikas Nandeshwar	“Study on Ethnobotanical Survey of Kurkheda tahshil”	Field Work, Survey	Mr. Amar S. Kuril
5.	Miss. Sapana Dubraj Meshram	“Seed Germination of Pulses”	Lab Work	Miss. Priyanka M. Sahare
6.	Miss. Ankisha Fakira Chaudhari	“Studies on plants used for purpose of fencing and agricultural implements by the local people of Gadchiroli tahshi.”	Field Work, Survey	Dr. P.S.Jakhi
7.	Miss. Mayuri waman Borkar	“Study of fruit marketing from Gadchiroli”	Field Work, Survey	Dr. Prashant S. Jakhi
8.	Miss. Raksha Satywan Raut	“Agricultural seeds , Pesticides & Fertilizer available in kurkheda tehsil.”	Field Work, Survey	Dr. P. S. Jakhi
9.	Miss. Ragini Machhindranath Lakade	“studies on Dye yielding plants from gadchiroli district of Maharashtra.”	Field Work, Survey	Mr. Amar S. Kuril
10	Miss. Sejal Vinod Sakhare Supervisor Name -	“Documentation of wild and cultivated vegetables in Gadchiroli tehsil”	Field Work, Survey	Miss. Kalyani G. Khobragade
11	Miss. Pallavi Shamrao Kadyami	“Aquatic plants diversity of Gadchiroli Tehsil”	Field Work, Survey	Mr. Amar S. Kuril
12	Miss. Achal Homraj	“Documentation on thorny,	Field Work,	

	Chahande	spiny and prickles plants of Gadchiroli Tahsil."	Survey	Miss. Kalyani G. Khobragade
13	Miss. Urvashi Gowardhan Uikey	"Biodiversity assessment of aquatic plants from few selected sites in Kurkheda tehsil, District Gadchiroli"	Field Work, Survey	Mr. Amar S. Kuril
14	Miss. Revata Sitaram Sondarkar Supervisor Name: Priyanka M. Sahare	"Leaf epidermal studies of some species of Apocynaceae and Asteraceae"	Lab Work	Miss. Priyanka M. Sahare
15	Miss. Mayuri kalidas Gurnule	"Checklist of plants available in premises of Government Science College, Gadchiroli."	Field Work, Survey	Dr. P. S. Jakhi
16	Miss. Babita Ramesh Buddhe.	"Qualitative studies on Agricultural diversity and their soil testing of few selected sites from Desaiganj tahsil dist.Gadchiroli"	Lab Work	Mr. Amar S. Kuril

Date: - 28/04/2024

  
Principal  
Govt. Science College  
Gadchiroli

## OEE Result for Academic Year 2024-2025

This is not the Merit List; it is only the result of the OEE 2024.

\* Search Options:



Program Name  
Wise



User Name  
Wise



Application ID  
Wise

\* Enter User Name / Email ID:

AKANKSHAPATLE74@GMAIL.COM

## Result

Category	Domicile	University	Divyanag	Total Marks
C	Maharashtra	Other University	No	<b>19.25</b>

For Technical queries mail us from your registered e-Mail address at - [CSPSupport@pun.unipune.ac.in](mailto:CSPSupport@pun.unipune.ac.in) OR call us at **020-71533899**



Applicant Name Akanksha  
Jitendra Patle

Status You have not submitted a valid Category Certificate. Until you upload a valid Category Certificate, you will be considered under GENERAL Category. The last date for submission of the Category Certificate is 31st March 2024.

Test Paper(s)	Marks Scored out of 100
Biotechnology (BT)	28.33

Government Science college Gadchiroli

DEPARTMENT OF BOTANY

"Internal assignment submission and  
seminar" "Anther culture"  
submitted by,

Name: Tabish M. Pathan

Class: Bsc. III yr, sem - VI

session: winter / summer - 2024 - ~~2025~~

submitted to,

Teachers incharge

1. Plant Biotech - I ~~Dr. P. S. Takhi~~  
06/02/2025

2. Plant Biotech - II ~~Dr. P. S. Takhi~~  
20/2/24

Seminars - Anther culture

~~Dr. P. S. Takhi~~  
20/2/24

~~Dr. P. S. Takhi~~  
Dr. P. S. Takhi

Head Department  
of Botany

### Q.1 Embryogenesis (somatic and zygotic)

An embryo is defined as a plant in its initial stage of development. Each embryo possesses two distinct poles, one to form root and the other shoot, and is the product of fusion of gametes. In some plant species, embryos are produced without the fusion of gametes and termed as asexual embryogenesis or adventitious embryony.

In an intact plants this type of embryogenesis may occur in sporophytic tissues like integuments, nucellar tissues or from unfertilized gametic cells. Apart from the normal cases of embryo formations such as, zygotic embryogenesis and adventitious embryony, instances of embryo formations from the tissues culture in vitro were reported. This phenomenon, termed as somatic embryogenesis was first observed by Steward and his co-workers (1958) in suspension cultures of carrot followed by Reinert (1959). Since then a number of reports of embryo formation have been published.



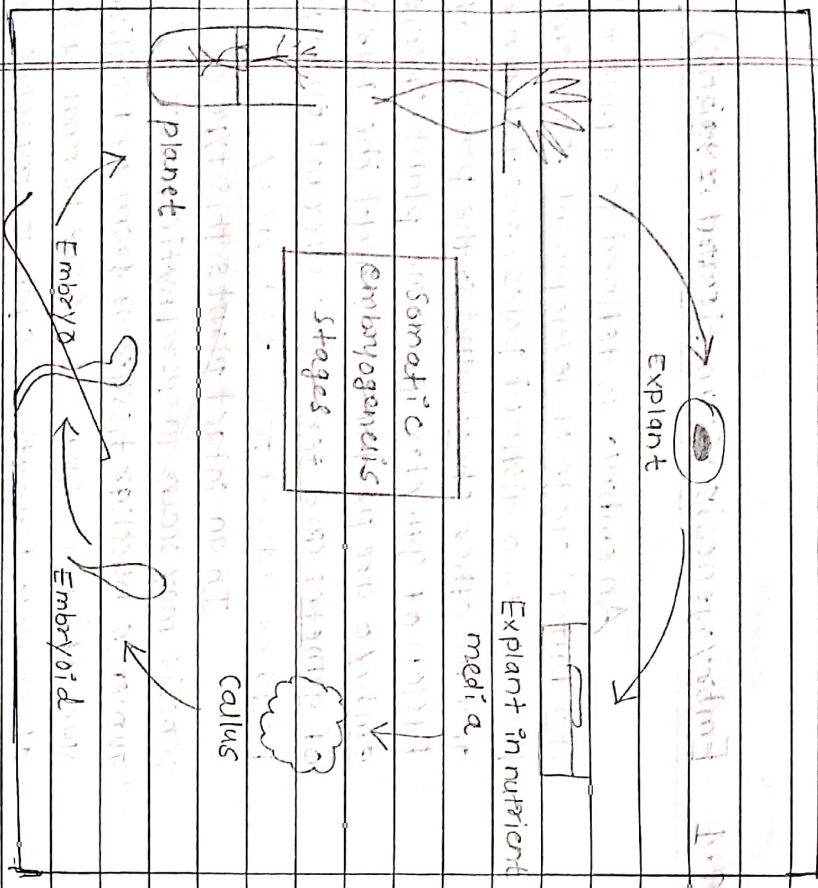


Fig. Somatic embryogenesis stages

Somatic embryogenesis in vitro produces embryos like structures resembling the zygotic embryos in structure and morphogenic potential. Despite this resemblance, the origin and development of an embryo-like structure from somatic cell differs from that of zygotic embryo, where the origin is from a single cell.

Embryoid is generally used to denote the embryo-like structures from cultured tissues.

Q.2

Major types of tissue culture media (MS, B5, NG)

1.

Murashige and Skoog medium (MS)

Murashige and Skoog (MS) originally formulated a medium in 1962, to induce organogenesis, and systems. Murashige & Skoog medium (MS) is used for micropropagation organ culture, callus culture and cell suspension culture. Murashige and Skoog medium (MS) is established by Murashige and Skoog (1962) for in vitro callus culture and Nicotiana glauca (family - Solanaceae). Murashige and Skoog Medium (MS) provides all essential macroelements and vitamins for the growth of plant cells, tissue and organ culture. In vitro. Medium with high concentration of salts is used for cultivating plant cells, tissue and organ culture.

2.

B5 medium

The B5 medium developed by O.L. Gamborg in 1968, was originally designed for cell suspension and callus cultures of Glycine max. At present with certain modifications

## Seminar

Despite being the first to develop golden rice in century, in 2017 a group of Indian researchers reported that genes needed to produce Golden Rice have uninucleated effects when they inserted the engineered DNA in the high yielding and agronomically superior Indian rice variety Soomra, it became pale and stunted. The yields were so reduced that it was unsuitable for cultivation. There has not been much progress since for development of golden rice in India.

20/11/2024

### \* Anther culture

Anther culture means plant regeneration from the haploid microspore cells with the aim of haploid and diploid plant.

Anther culture was first reported in the 1970s through in methods by Chen and Maheshwari. from the plant batata.

Anther culture is a technique by which the technique developing anthers at a precise and critical stage are excised aseptically from unopened flower bud and are cultured in a nutrient medium where the microspores with cultured anthers develop into callus tissue or embryoids that give rise to haploid plantlets either through organogenesis or embryogenesis.

### • Pollen culture

Pollen or microspore culture is an in vitro technique by which the pollen grains preferably at the uninucleated stage, are squeezed out aseptically from the intact anthers and then cultured on nutrient medium. The microspores develop into embryoids or callus tissue that gives to rise plantlets by embryogenesis or organogenesis.

• Anther culture

The selected flower buds of young plants are surface sterilized and anthers removed along with their filaments. The anthers are excised under aseptic conditions and crushed in 1% acetocarmine to test the stage of pollen development.

If they are at the correct stage, each anther is gently separated (from the filament) and the intact anthers are inoculated on a nutrient medium. Injured anthers should not be used in cultures as they result in callusing of anther wall tissue.

The anther cultures are maintained in alternating period of light (12 hrs) and darkness (6-12 hrs) at 28°C. As the anthers proliferate they produce callus which later forms an embryo and then a haploid plant.

\* Androgenesis:

Anther/pollen culture is referred androgenesis (the male gametophyte microspore or immature pollen produces haploid plant). The cultured microspores mainly follow four distinct pathways during initial stages of in vitro androgenesis:

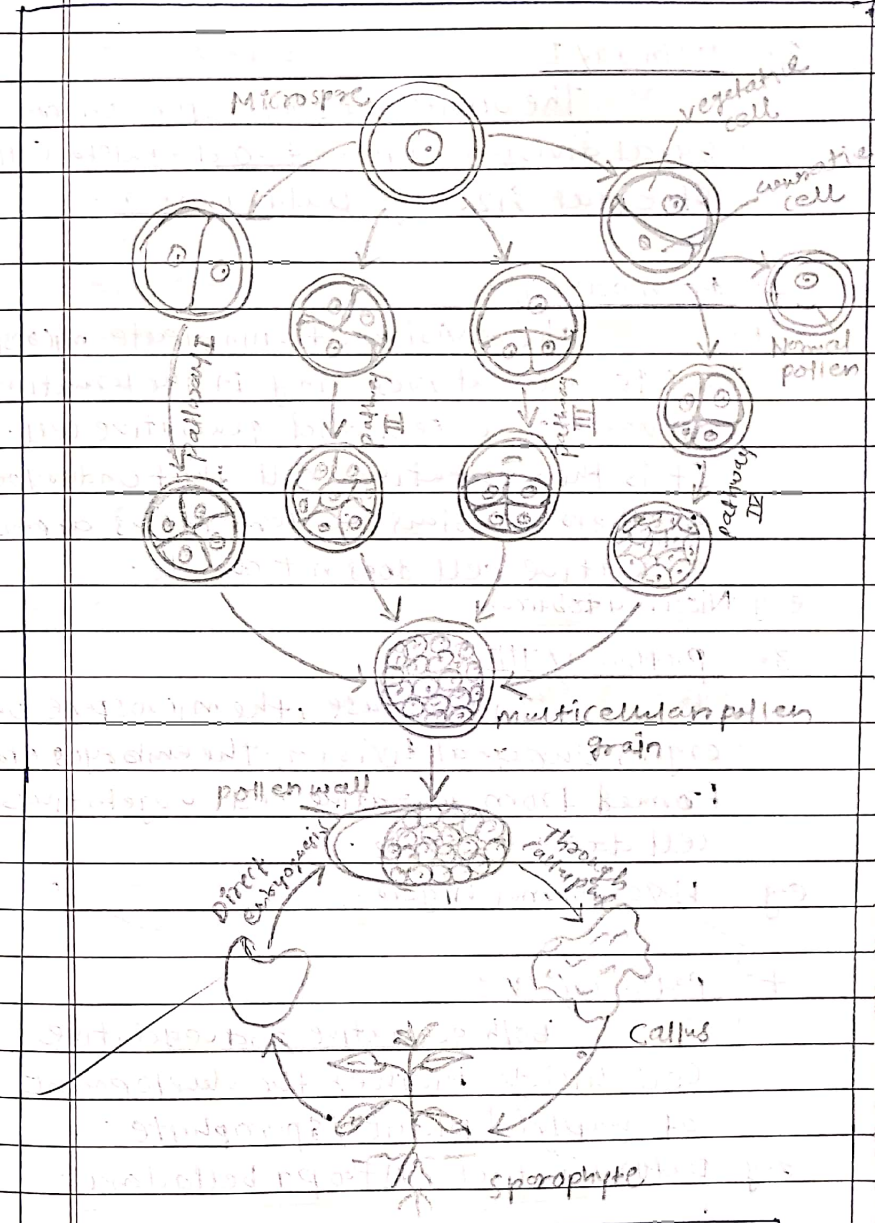


Fig. Diagram showing the origin of sporophytes from pollen grains in anther culture.

1. Pathway I:

The uninucleate microspore undergoes equal division to form two daughter cells of equal size e.g. Datura innoxia.

2. Pathway II:

The division of uninucleate microspores is unusual, resulting in the formation of vegetative cell and generative cell. It is the vegetative cell that undergoes further divisions to form callus or embryo. Generative cell does not divide.

e.g. Nicotina tobaccum3. Pathway III:

In this case, the microspore undergoes unequal division. The embryos are formed from generative cell. Vegetative cell does not divide.

e.g. Hyoscyamus niger.4. Pathway IV:

Both generative and vegetative cell divide further the development of haploid plant / sporophyte.

e.g. Datura met al, Atropa belladana.

At the initial stages, the microspore may follow any one of the four pathways described above. As the cell divide, the pollen grain becomes multicellular and burst open. This multicellular mass may form a callus which later differentiates into a plant (through callus phase). Alternatively the multicellular mass may produce the plant through direct embryogenesis.

\* Advantages of Anther Culture

- simple technique
- Less time consuming
- A high frequency of haploid plants, which is easily identified by their smaller sterile flowers.
- Easy to induce cell division in most species.
- No requirement for very high level of expertise.